Biomarkers for Autism Spectrum Disorders (ASD): A Meta-analysis

Ashley Ansel, M.Sc.¹, Yehudit Posen, Ph.D.^{1,2}, Ronald Ellis, Ph.D.^{1,3}, Lisa Deutsch, Ph.D.⁴, Philip D. Zisman, Ph.D.¹, and Benjamin Gesundheit, M.D., Ph.D.^{1*}

¹Cell-El Therapeutics Ltd, Jerusalem, Israel; ²PSW Ltd, Rehovot, Israel; ³Biotech & Biopharma Consulting, Jerusalem, Israel; and ⁴Biostats Statistical Consulting Ltd, Modiin, Israel

ABSTRACT

Objective: To compare the reported accuracy and sensitivity of the various modalities used to diagnose autism spectrum disorders (ASD) in efforts to help focus further biomarker research on the most promising methods for early diagnosis.

Abbreviations: ADI-R, Autism Diagnostic Interview–Revised; ADOS, Autism Diagnostic Observation Schedule; ASD, autism spectrum disorders; AUC, area under the curve; CARS, Childhood Autism Rating Scale; DSM, Diagnostic and Statistical Manual of Mental Disorders; EEG, electroencephalography; ELISA, enzyme-linked immunosorbent assay; fMRI, functional magnetic resonance imaging; HPLC, high-performance liquid chromatography; LC-HRMS, liquid chromatography–high resolution mass spectrometry; MRI, magnetic resonance imaging; NMR, nuclear magnetic resonance; PDD, pervasive developmental disorders; PCR, polymerase chain reaction; PET, positron emission tomography; PRISMA, Preferred Reporting Items for Systematic Reviews and Meta-Analyses; RRB, restricted repetitive behaviors; rs-fMRI, resting state functional magnetic resonance imaging; sMRI, structural magnetic resonance imaging; SVM, support vector machine; TD, typically developing; UV-Vis, ultraviolet/visible spectroscopy; VBM, voxel-based morphometry.

Citation: Ansel A, Posen Y, Ellis R, Deutsch L, Zisman PD, Gesundheit B. Biomarkers for Autism Spectrum Disorders (ASD): A Meta-analysis. Rambam Maimonides Med J 2019;10 (4):e0021. doi:10.5041/RMMJ.10375

Copyright: © 2019 Ansel et al. This is an open-access article. All its content, *except where otherwise noted*, is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Acknowledgements: The authors thank Joshua Rosenzweig for reviewing the literature, Frederic and Netanel Deutsch for assisting in the statistical analysis. The study was supported by Cell-El Ltd, a company seeking to define ASD-specific biomarker signatures to enable objective, biomarker-based ASD diagnostics.

Author contributions statement: AA reviewed the relevant literature, made substantial contributions to conception and design, and interpreted the data. YP reviewed the relevant literature and assisted in drafting the article. RE interpreted the findings and participated in revising the article and approving the final version. LD performed the statistical analyses. PDZ made substantial contributions to conception and design and participated in interpreting the data and in revising the paper. BG conceived the performing of the meta-analysis, developed its original detailed plan, participated in revising the paper, and approved the final version. All authors approved the final manuscript as submitted and agree to be accountable for all aspects of the work.

Conflict of interest: AA, PDZ, and BG are employees of Cell-El Ltd, a company seeking to identify autism-specific biomarkers. YP, RE, and LD serve as consultants at Cell-El Ltd. Cell-El Ltd had no control over the study data, analysis, interpretation, or the decision to publish.

* To whom correspondence should be addressed. E-mail: GesundheitMD@cell-el.com

Methods: The Medline scientific literature database was searched to identify publications assessing potential clinical ASD biomarkers. Reports were categorized by the modality used to assess the putative markers, including protein, genetic, metabolic, or objective imaging methods. The reported sensitivity, specificity, area under the curve, and overall agreement were summarized and analyzed to determine weighted averages for each diagnostic modality. Heterogeneity was measured using the I^2 test.

Results: Of the 71 papers included in this analysis, each belonging to one of five modalities, protein-based followed by metabolite-based markers provided the highest diagnostic accuracy, each with a pooled overall agreement of 83.3% and respective weighted area under the curve (AUC) of 89.5% and 88.3%. Sensitivity provided by protein markers was highest (85.5%), while metabolic (85.9%) and protein markers (84.7%) had the highest specificity. Other modalities showed degrees of sensitivity, specificity, and overall agreements in the range of 73%–80%.

Conclusions: Each modality provided for diagnostic accuracy and specificity similar or slightly higher than those reported for the gold-standard Autism Diagnostic Observation Schedule (ADOS) instrument. Further studies are required to identify the most predictive markers within each modality and to evaluate biological pathways or clustering with possible etiological relevance. Analyses will also be necessary to determine the potential of these novel biomarkers in diagnosing pediatric patients, thereby enabling early intervention.

KEY WORDS: Autism spectrum disorder, biomarkers, gene expression, magnetic resonance imaging, meta-analysis, proteomics

INTRODUCTION

Autism spectrum disorders (ASD) were first characterized clinically in 1943 by Kanner¹ and further in 1979 by Wing and Gould² as a spectrum of impaired social interactions, restricted communications skills, and unusual repetitive behaviors. The American Psychological Association's *Diagnostic and Statistical Manual of Mental Disorders, Fifth Edition* (DSM-5) recently consolidated the various subtypes of pervasive developmental disorders (PDD) into one category called ASD and shifted the evaluation from three domains (social deficits, communication deficits, and restricted repetitive behaviors [RRB]) to two (social-communication impairments and RRB).³

Several behavior assessment-based diagnostic tests have been used for ASD, including the Autism Diagnostic Observation Schedule (ADOS).⁴ A metaanalysis involving ADOS evaluations of >4,000 children reported an overall diagnostic accuracy of 52%, with sensitivity scores of 67%–97% (pooled data: 91%) and specificity scores of 56%–94% (pooled data: 73%).⁵ The Autism Diagnostic Interview– Revised (ADI-R) is a structured interview of the parent, differing from the direct observation of the child performed with the ADOS evaluation.⁶ In addition, clinicians use the Childhood Autism Rating Scale (CARS) to rate the child's behavior on 15 subscales;⁷ parental reports can also be considered.

Since these behavioral tests are subjective and time-consuming, require professional staff to be administered, and can only be used from age 3 years once the child is old enough to communicate, researchers have sought other ways of diagnosing ASD.8 Biomarkers are expected to be more objective, should enable earlier diagnosis, and may provide clues to the underlying etiology of ASD. In addition, providing positive diagnosis in younger toddlers may enable earlier initiation of therapy with consequently higher probability of successful treatment given decreasing brain plasticity with age in the developing child. The primary modalities harnessed to identify novel ASD biomarkers have been molecular, proteomic, metabolomic, neurochemical, radiologic, and electrophysiologic, with transcriptomic analyses also having been performed.9

This study aimed to compare the reported accuracy and sensitivity of the various modalities used to diagnose ASD. This should help focus further biomarker research on the most promising methods for early diagnosis.

METHODS

This analysis aimed to adhere to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines, to ensure comprehensive and transparent reporting.

Search Strategy

Independent queries using medical subject headings (MeSH) and keywords were performed to identify all primary research articles from the PubMed database that evaluated sensitivity, specificity, and accuracy of biomarkers for diagnosing ASD within six predefined modalities (protein, metabolic, genetic, electroencephalography [EEG], magnetic resonance imaging [MRI], positron emission tomography [PET]) (Supplement). Each search term included autism/ASD, a diagnostic technique, and a sensitivity/specificity classifier. To distinguish between protein and metabolic papers, all papers dealing with hormones, urine, mass spectrophotometry, metabolites, or peptides were assigned to the metabolic modality, while all papers dealing with cytokines, chemokines, or other proteins circulating in the blood were assigned to the protein modality.

Data Sources and Data Extraction

The search terms were entered into Ovid MEDLINE (1946 to January 2017) without limits, and 866 articles were returned. Reviews and reference lists were cross-checked for studies that the search terms might have missed.

Screening

Two independent reviewers (AA and JR) examined study titles. From review of the abstracts, potentially eligible full-text articles were retrieved with relevant appendices and supplementary information.

Eligibility

Full-text articles were reviewed against eligibility criteria. Inclusion criteria were: (a) articles after 1994, written in English; (b) inclusion of a typically developing (TD) control group (unless it was a review paper); (c) inclusion of a study group with children diagnosed with ASD by a behavioral diagnostic test or by DSM criteria (unless it was a review paper); and (d) assessment for ASD biomarkers using one of the six predefined modalities. Exclusion criteria were: (a) studies that compared ASD with other comorbidities; (b) studies that tested only risk factors of ASD; (c) studies that had a therapeutic component; (d) non-clinical studies; and (e) studies without statistical parameters of interest (sensitivity, specificity, accuracy, and/or area under the curve [AUC]). All publications meeting all inclusion criteria and none of the exclusion criteria were included in the analysis.

Data Extraction

For each eligible article, the following data were extracted and validated independently by two researchers (AA and JR): first author's surname, year of publication, diagnostic modality, number of ASD subjects, number of controls, age-matched (yes or no), sex-matched (yes or no), accuracy (% correctly identified), AUC, sensitivity, and specificity. The standard error of the AUC was calculated based on the AUC point estimate and sample size by the method of Hanley and McNeil.¹⁰ Several papers included the evaluation of multiple markers assessed on the same group of subjects; these were included in the meta-analysis individually and analyzed independently of each other.

Data Analysis

A separate statistical analysis of each of the parameters was performed for each of the six predefined modalities. Meta-analyses of sensitivity, specificity, AUC, and accuracy were performed using MedCalc Statistical Software version 17.9.7 (MedCalc Software bvba, Ostend, Belgium). The weighted summary sensitivity, specificity, accuracy (arc-sine square root transformation), and AUC with 95% confidence intervals (CIs) were calculated using a random-effects model.¹¹ Inter-study heterogeneity was assessed using the I^2 statistic with 95% CI, which describes the percent variability in point estimate due to heterogeneity rather than sampling error. Presented are the weighted summaries of each tested parameter, alongside the weighted summary of the ADOS test, as reported in a published meta-analysis.⁵ The contribution of each individual publication to the weighted summary of each measured parameter is presented by modality.

RESULTS

The literature search identified 866 papers (Figure 1), of which 211 were duplicates. The abstracts of the remaining 655 papers were screened for relevance; 29 were excluded. Thus, 626 full-text papers were assessed for eligibility based on the above criteria. Review of these papers identified 86 additional papers which were also further appraised. Of these 712 publications in total, 640 failed to meet the inclusion criteria, primarily due to lack of an ASD group or insufficient statistical data. The 72 papers that met the criteria were subdivided by the diagnostic modality reported in the study. Four papers included data belonging to two diagnostic



Figure 1. PRISMA Flow Diagram of the Phases of the Literature Search.

modalities.^{12–15} Since only one PET study met the eligibility criteria, this modality was excluded from the analysis. Cohort sizes varied from 6 to 554 subjects.

Genetic studies $(n=14)^{1_3,14,16-27}$ applied polymerase chain reaction (PCR) genotyping, mRNA/miRNA microarray, or spectrophotometric tools. Magnetic resonance imaging (MRI) studies $(n=22),^{1_3,14,21,28-46}$ which included functional MRI (fMRI), resting state fMRI (rs-fMRI), structural MRI (sMRI), and standard MRI studies, sought out ASD diagnostic markers by analyzing fast spin echo (FSE) T2-weighted, fluid-attenuated inversion recovery (FLAIR),

diffusion-weighted imaging (DWI), spin echo (SE) T1-weighted sequences, and single voxel 1H MR spectrum. Volumetric measurements for different areas of the brain were assessed. Mass univariate methods such as voxel-based morphometry (VBM) and whole-brain classification approach employing a support vector machine (SVM) were used. In the 21 metabolic studies, 12, 15, 47-65 high-performance liquid chromatography (HPLC), liquid chromatographyhigh resolution mass spectrometry (LC-HRMS), gas chromatography, nuclear magnetic resonance (NMR), as well as capillary electrophoresis with ultraviolet/ visible spectroscopy (UV-Vis) were used to identify markers. The 12 studies assessing the diagnostic potential of various protein markers12,15,66-75 covered >60 proteins or protein combinations using various microarray kits/chips or enzyme-linked immunosorbent assay (ELISA). Six studies focused on EEG analyses for ASD diagnosis.76-81

Overall, protein-based followed by metabolitebased studies provided the highest diagnostic accuracy, each with an overall agreement of 83.3% and AUC of sensitivity (true-positive rates) versus specificity (false-positive rates) of 89.5% and 88.3%, respectively (Table 1, Figures 2–5). Sensitivity provided by protein markers (85.5%) and metabolite markers (84.7%) was highest. The other modalities showed similar degrees of sensitivity, specificity, and overall agreements, which all fell within the range of 73%–80%.

DISCUSSION

As the first reported meta-analysis of ASD biomarkers, the current study included 71 papers with cohorts of up to 554 ASD subjects.⁷⁷ This study suggests that all five major bio-diagnostic modalities provide similar diagnostic objective accuracy for ASD compared to the subjective ADOS. The proteinand metabolite-based tests were found to provide for the highest diagnostic accuracy; combining modalities might further improve diagnostic accuracy.

The sensitivities of the various studied modalities were 73.6%–85.5%, with protein markers showing the highest degree of sensitivity. The specificities ranged from 73.0% (MRI) to 85.9% (metabolic). Accuracy, assessed by overall agreement and AUC, was 73.4%–83.2% and 79.0%–89.5%, respectively, and highest for protein-based biomarkers. Taken together, all analyzed modalities provided for higher diagnostic accuracy and specificity compared to the gold-standard ADOS test.⁵ While pooled ADOS

Modality	Sensitivity (%)	Specificity (%)	Overall Agreement (%)	Area under Curve (AUC) (%)
Genetic				
Pool	79.3	73.1	76.7	79.5
Range	24.3-100	41.8-88.2	62.7-90.4	64.8-92.0
95% CI	73.3-84.7	69.6-76.5	73.8-79.5	77.0-82.0
l ²	87.5	56.9	70.8	52.5
MRI				
Pool	73.6	73.0	73.5	79
Range	43.7-94.8	45.4-100	57.9-95.7	58.0-99.5
95% CI	71.8-76.0	70.0-76.0	71.0-75.9	75.0-83.0
l ²	78.1	84.4	89.1	96.4
Metabolic				
Pool	74.6	85.9	83.3	88.3
Range	0-100	15.0-100	48.7-100	59.2-99.9
95% CI	66.8-81.6	82.7-88.7	80.3-86.1	86.0-91.0
l ²	97.7	87.1	ND	92.2
Protein				
Pool	85.5	84.7	83.8	89.5
Range	50-100	38.9-100	62.8-100	57.0-99.9
95% CI	80.0-90.3	78.6-90	81.0-86.5	86.0-93.0
l ²	87.3	89.2	73	90.1
EEG				
Pool	79.9	80.4	79.9	ND
Range	57.6-90.9	64.5-100	70.5-87.4	ND
95% CI	70.5-87.9	73.3-86.6	73.5-85.5	ND
l ²	83.4	79.5	85.2	ND

Table 1. Weighted ASD Diagnostic Power for Each Evaluated Modality.

ND, not done.

diagnostic sensitivity was higher than for biomarker modalities, protein-based diagnoses provided for sensitivity within the same range. This metaanalysis supports efforts to search for/use new objective modalities beyond psychological tests for the diagnosis of ASD. Moreover, quantitative objective biomarkers identified at ages when psychological tests cannot yet be employed should enable earlierstage intervention, which is projected to be more efficient due to greater brain plasticity. These diagnostic efforts may enable the subdivision of ASD into subgroups and provide useful therapeutic targets, which have significant long-term therapeutic implications.

Further studies will be necessary to determine which modalities serve better as screening versus confirmatory testing. Subgroups of ASD might be defined by these tests, suggesting different therapeutic modalities as diagnostic targets for different

		Sensitivity	Lower 95% Confidence Limit	Upper 95% Confidence Limit
EEG	⊢ • 1	79.91%	70.45%	87.94%
Genetic	II	79.29%	73.31%	84.69%
MRI	⊢ •	73.64%	71.18%	76.04%
Metabolic	·	74.59%	66.84%	81.62%
Protein	 1	85.53%	80.02%	90.29%
ADOS		91.00%	89.00%	93.00%
	70 80 90	100		



Figure 2. Weighted Sensitivity of Appraised Studies.

The weighted sensitivity with 95% CIs was calculated using a random-effects model. Also shown is the weighted sensitivity of the ADOS test, as determined in a meta-analysis of seven cross-sectional studies assessing >4,000 children.⁵

		Specificity	Lower 95% Confidence Limit	Upper 95% Confidence Limit
EEG	┝━━┥	80.37%	73.25%	86.63%
Genetic	H=-1	73.12%	69.60%	76.50%
MRI	⊢ - -1	73.03%	70.01%	75.95%
Metabolic	┝┻┥	85.85%	82.74%	88.70%
Protein	⊢ •-1	84.74%	78.62%	89.98%
ADOS ++		27.00%	24.00%	31.00%
20 40	60 80 1	00		



Figure 3. Weighted Specificity of Appraised Studies.

The weighted specificity with 95% CIs was calculated using a random-effects model. Also shown is the weighted sensitivity of the ADOS test, as determined in a meta-analysis of seven cross-sectional studies assessing >4,000 children.⁵

	Overall Agreement	Lower 95% Confidence Limit	Upper 95% Confidence Limit
EEG H	79.85%	73.53%	85.51%
Genetic ⊣—	76.71%	73.76%	79.54%
MRI ————————————————————————————————————	73.47%	70.97%	75.88%
Metabolic Hand	83.32%	80.31%	86.13%
Protein	83.82%	80.99%	86.46%
70 75 80 85 90 95 10	00		
Overall Agreement (%)			

Figure 4. Weighted Overall Agreement of Appraised Studies.

The weighted overall agreement with 95% CIs was calculated using a random-effects model.

			Sensitivity	Lower 95% Confidence Limit	Upper 95% Confidence Limit
EEG	⊦∎		79.91%	70.45%	87.94%
Genetic	·•		79.29%	73.31%	84.69%
MRI	⊢ ∎–-		73.64%	71.18%	76.04%
Metabolic	·•	4	74.59%	66.84%	81.62%
Protein	F		85.53%	80.02%	90.29%
ADOS			91.00%	89.00%	93.00%
	70 80	90	100		
	Sens	itivity (%)			



The weighted AUC with 95% CIs were calculated using a random-effects model.

such subgroups. Attempts to find patterns linking the most accurate biomarkers in each modality may identify common pathways and draw the ASD community closer to developing therapeutics, where these diagnostic markers will serve with psychological tests as objective theragnostic monitoring tools.

There were several limitations in this metaanalysis. There was a high level of heterogeneity, as expected given both the clinical heterogeneity between the included papers, with variations in the diagnosis and definition of ASD, and the methodologies across studies. In addition, in all assessed publications, evaluation of the diagnostic value of the biomarker of interest used typically developing controls as a comparator group, which may falsely elevate the diagnostic capacity of the test as compared to its performance in marginal cases with behaviors consistent with ASD. Comparing across modalities was not uniformly well-controlled. In addition, certain modalities were documented in a very limited number of papers. Furthermore, many papers were excluded due to insufficient statistical data, reinforcing the importance of proper study design and execution in future analyses of ASD biomarkers.

In conclusion, this study suggests that five major bio-diagnostic modalities provide a higher level of accuracy for objective diagnosis of ASD compared to ADOS, the gold-standard test. Thus, it is justified to include objective biological tests in the diagnosis of ASD to develop and monitor future biological therapies. Further studies looking at each modality in higher resolution to fine-tune the findings are still necessary. Objective biomarkers together with current psychological evaluations might enable improved diagnosis and monitoring.

REFERENCES

- 1. Kanner L. Autistic disturbances of affective contact. Acta Paedopsychiatr 1968;35:100–36.
- 2. Wing L, Gould J. Severe impairments of social interaction and associated abnormalities in children: epidemiology and classification. J Autism Dev Disord 1979;9:11–29.
- 3. Lord C, Bishop SL. Recent advances in autism research as reflected in DSM-5 criteria for autism spectrum disorder. Annu Rev Clin Psychol 2015;11: 53–70. <u>Crossref</u>
- 4. Lord C, Rutter M, Goode S, et al. Autism diagnostic observation schedule: a standardized observation of

communicative and social behavior. J Autism Dev Disord 1989;19:185–212.

- 5. Tsheringla S, Minju KA, Russell S, Mammen P, Russell PS, Nair MK. A meta-analysis of the diagnostic accuracy of Autism Diagnostic Observation Schedule Module-1 for autism spectrum disorders. Indian J Pediatr 2014;81(Suppl 2):S187–92. <u>Crossref</u>
- Lord C, Rutter M, Le Couteur A. Autism Diagnostic Interview-Revised: a revised version of a diagnostic interview for caregivers of individuals with possible pervasive developmental disorders. J Autism Dev Disord 1994;24:659–85.
- Schopler E, Reichler RJ, DeVellis RF, Daly K. Toward objective classification of childhood autism: Childhood Autism Rating Scale (CARS). J Autism Dev Disord 1980;10:91–103.
- 8. Gesundheit B, Rosenzweig JP. Editorial: Autism spectrum disorders (ASD)—searching for the biological basis for behavioral symptoms and new therapeutic targets. Front Neurosci 2017;10:607. Crossref
- 9. Ansel A, Rosenzweig JP, Zisman PD, Melamed M, Gesundheit B. Variation in gene expression in autism spectrum disorders: an extensive review of transcriptomic studies. Front Neurosci 2017;10:601. <u>Crossref</u>
- 10. Hanley JA, McNeil BJ. The meaning and use of the area under a receiver operating characteristic (ROC) curve. Radiology 1982;143:29–36. <u>Crossref</u>
- 11. DerSimonian R, Laird N. Meta-analysis in clinical trials. Control Clin Trials 1986;7:177–88.
- El-Ansary A, Al-Ayadhi L. Neuroinflammation in autism spectrum disorders. J Neuroinflammation 2012;9:265. <u>Crossref</u>
- 13. Iidaka T. Resting state functional magnetic resonance imaging and neural network classified autism and control. Cortex 2015;63:55–67. <u>Crossref</u>
- Kang DW, Park JG, Ilhan ZE, et al. Reduced incidence of Prevotella and other fermenters in intestinal microflora of autistic children. PLoS One 2013;8: e68322. <u>Crossref</u>
- 15. Yang CJ, Liu CL, Sang B, Zhu XM, Du YJ. The combined role of serotonin and interleukin-6 as biomarker for autism. Neuroscience 2015;284:290–6. <u>Crossref</u>
- Croen LA, Goines P, Braunschweig D, et al. Brainderived neurotrophic factor and autism: maternal and infant peripheral blood levels in the Early Markers for Autism (EMA) Study. Autism Res 2008; 1:130–7. Crossref
- 17. Gaita L, Manzi B, Sacco R, et al. Decreased serum arylesterase activity in autism spectrum disorders. Psychiatry Res 2010;180:105–13. <u>Crossref</u>

- Glatt SJ, Tsuang MT, Winn M, et al. Blood-based gene expression signatures of infants and toddlers with autism. J Am Acad Child Adolesc Psychiatry 2012;51:934–44.e2. <u>Crossref</u>
- Guan J, Yang E, Yang J, Zeng Y, Ji G, Cai JJ. Exploiting aberrant mRNA expression in autism for gene discovery and diagnosis. Hum Genet 2016; 135:797–811. Crossref
- 20. Hicks SD, Ignacio C, Gentile K, Middleton FA. Salivary miRNA profiles identify children with autism spectrum disorder, correlate with adaptive behavior, and implicate ASD candidate genes involved in neurodevelopment. BMC Pediatr 2016;16:52. <u>Crossref</u>
- Katuwal GJ, Cahill ND, Baum SA, Michael AM. The predictive power of structural MRI in autism diagnosis. Conf Proc IEEE Eng Med Biol Soc 2015; 2015:4270-3. <u>Crossref</u>
- 22. Latkowski T, Osowski S. Computerized system for recognition of autism on the basis of gene expression microarray data. Comput Biol Med 2015;56:82–8. <u>Crossref</u>
- 23. Maekawa M, Yamada K, Toyoshima M, et al. Utility of scalp hair follicles as a novel source of biomarker genes for psychiatric illnesses. Biol Psychiatry 2015; 78:116–25. <u>Crossref</u>
- 24. Meng WD, Sun SJ, Yang J, Chu RX, Tu W, Liu Q. Elevated serum brain-derived neurotrophic factor (BDNF) but not BDNF gene Val66Met polymorphism is associated with autism spectrum disorders. Mol Neurobiol 2016;54:1167–72. <u>Crossref</u>
- 25. Mundalil Vasu M, Anitha A, Thanseem I, et al. Serum microRNA profiles in children with autism. Mol Autism 2014;5:40. <u>Crossref</u>
- 26. Oh DH, Kim IB, Kim SH, Ahn DH. Predicting autism spectrum disorder using blood-based gene expression signatures and machine learning. Clin Psychopharmacol Neurosci 2017;15:47–52. <u>Crossref</u>
- 27. Pramparo T, Pierce K, Lombardo MV, et al. Prediction of autism by translation and immune/inflammation coexpressed genes in toddlers from pediatric community practices. JAMA Psychiatry 2015;72: 386–94. <u>Crossref</u>
- Anderson JS, Nielsen JA, Froehlich AL, et al. Functional connectivity magnetic resonance imaging classification of autism. Brain 2011;134(Pt 12):3742–54. Crossref
- 29. Casanova MF. Neuropathological and genetic findings in autism: the significance of a putative minicolumnopathy. Neuroscientist 2006;12:435–41. Crossref
- 30. Chen CP, Keown CL, Jahedi A, et al. Diagnostic classification of intrinsic functional connectivity high-

lights somatosensory, default mode, and visual regions in autism. Neuroimage Clin 2015;8:238–45. <u>Crossref</u>

- 31. Ecker C, Rocha-Rego V, Johnston P, et al. Investigating the predictive value of whole-brain structural MR scans in autism: a pattern classification approach. Neuroimage 2010;49:44–56. <u>Crossref</u>
- 32. Ghiassian S, Greiner R, Jin P, Brown MR. Using functional or structural magnetic resonance images and personal characteristic data to identify ADHD and autism. PLoS One 2016;11:e0166934. <u>Crossref</u>
- 33. Gori I, Giuliano A, Muratori F, et al. Gray matter alterations in young children with autism spectrum disorders: comparing morphometry at the voxel and regional level. J Neuroimaging 2015;25:866–74. <u>Crossref</u>
- Ingalhalikar M, Kanterakis S, Gur R, Roberts TP, Verma R. DTI based diagnostic prediction of a disease via pattern classification. Med Image Comput Comput Assist Interv 2010;13(Pt 1):558–65. <u>Crossref</u>
- 35. Ingalhalikar M, Parker D, Bloy L, Roberts TP, Verma R. Diffusion based abnormality markers of pathology: toward learned diagnostic prediction of ASD. Neuroimage 2011;57:918–27. <u>Crossref</u>
- 36. Jiao Y, Chen R, Ke X, Chu K, Lu Z, Herskovits EH. Predictive models of autism spectrum disorder based on brain regional cortical thickness. Neuroimage 2010;50:589–99. <u>Crossref</u>
- Lange N, Dubray MB, Lee JE, et al. Atypical diffusion tensor hemispheric asymmetry in autism. Autism Res 2010;3:350–8. <u>Crossref</u>
- 38. Lim L, Marquand A, Cubillo AA, et al. Disorderspecific predictive classification of adolescents with attention deficit hyperactivity disorder (ADHD) relative to autism using structural magnetic resonance imaging. PLoS One 2013;8:e63660. <u>Crossref</u>
- Nielsen JA, Zielinski BA, Fletcher PT, et al. Multisite functional connectivity MRI classification of autism: ABIDE results. Front Hum Neurosci 2013;7:599. Crossref
- Plitt M, Barnes KA, Martin A. Functional connectivity classification of autism identifies highly predictive brain features but falls short of biomarker standards. Neuroimage Clin 2014;7:359–66. <u>Crossref</u>
- Shen MD, Nordahl CW, Young GS, et al. Early brain enlargement and elevated extra-axial fluid in infants who develop autism spectrum disorder. Brain 2013; 136(Pt 9):2825–35. <u>Crossref</u>
- 42. Uddin LQ, Supekar K, Lynch CJ, et al. Salience network-based classification and prediction of symptom severity in children with autism. JAMA Psychiatry 2013;70:869–79. <u>Crossref</u>

- 43. Varol E, Gaonkar B, Erus G, Schultz R, Davatzikos C. Feature ranking based nested support vector machine ensemble for medical image classification. Proc IEEE Int Symp Biomed Imaging 2012:146–9. <u>Crossref</u>
- 44. Wang L, Wee CY, Tang X, Yap PT, Shen D. Multi-task feature selection via supervised canonical graph matching for diagnosis of autism spectrum disorder. Brain Imaging Behav 2016;10:33–40. <u>Crossref</u>
- 45. Wee CY, Wang L, Shi F, Yap PT, Shen D. Diagnosis of autism spectrum disorders using regional and interregional morphological features. Hum Brain Mapp 2014;35:3414–30.
- Xiao X, Fang H, Wu J, et al. Diagnostic model generated by MRI-derived brain features in toddlers with autism spectrum disorder. Autism Res 2017;10: 620–30. <u>Crossref</u>
- Alabdali A, Al-Ayadhi L, El-Ansary A. Association of social and cognitive impairment and biomarkers in autism spectrum disorders. J Neuroinflammation 2014;11:4. <u>Crossref</u>
- Al-Yafee YA, Al-Ayadhi LY, Haq SH, El-Ansary AK. Novel metabolic biomarkers related to sulfurdependent detoxification pathways in autistic patients of Saudi Arabia. BMC Neurol 2011;11:139. <u>Crossref</u>
- 49. El-Ansary A, Al-Ayadhi L. Lipid mediators in plasma of autism spectrum disorders. Lipids Health Dis 2012;11:160. <u>Crossref</u>
- 50. El-Ansary AK, Bacha AG, Al-Ayahdi LY. Impaired plasma phospholipids and relative amounts of essential polyunsaturated fatty acids in autistic patients from Saudi Arabia. Lipids Health Dis 2011;10:63. <u>Crossref</u>
- 51. El-Ansary AK, Bacha AG, Al-Ayahdi LY. Plasma fatty acids as diagnostic markers in autistic patients from Saudi Arabia. Lipids Health Dis 2011;10:62. <u>Crossref</u>
- 52. Gevi F, Zolla L, Gabriele S, Persico AM. Urinary metabolomics of young Italian autistic children supports abnormal tryptophan and purine metabolism. Mol Autism 2016;7:47. <u>Crossref</u>
- 53. Heyer NJ, Echeverria D, Woods JS. Disordered porphyrin metabolism: a potential biological marker for autism risk assessment. Autism Res 2012;5:84–92. <u>Crossref</u>
- 54. Kuwabara H, Yamasue H, Koike S, et al. Altered metabolites in the plasma of autism spectrum disorder: a capillary electrophoresis time-of-flight mass spectroscopy study. PLoS One 2013;8:e73814. <u>Crossref</u>
- Li SO, Wang JL, Bjørklund G, Zhao WN, Yin CH. Serum copper and zinc levels in individuals with autism spectrum disorders. Neuroreport 2014;25:1216– 20. <u>Crossref</u>

- 56. Momeni N, Bergquist J, Brudin L, et al. A novel bloodbased biomarker for detection of autism spectrum disorders. Transl Psychiatry 2012;2:e91. <u>Crossref</u>
- 57. Nadal-Desbarats L, Aïdoud N, Emond P, et al. Combined 1H-NMR and 1H-13C HSQC-NMR to improve urinary screening in autism spectrum disorders. Analyst 2014;139:3460–8. <u>Crossref</u>
- 58. Shimmura C, Suda S, Tsuchiya KJ, et al. Alteration of plasma glutamate and glutamine levels in children with high-functioning autism. PLoS One 2011;6: e25340. <u>Crossref</u>
- 59. Soria AC, Wright B, Goodall DM, Wilson J. Data processing in metabolic fingerprinting by CE-UV: application to urine samples from autistic children. Electrophoresis 2007;28:950–64. <u>Crossref</u>
- 60. Testa C, Nuti F, Hayek J, et al. Di-(2-ethylhexyl) phthalate and autism spectrum disorders. ASN Neuro 2012;4:223–9. <u>Crossref</u>
- Vargason T, Howsmon DP, Melnyk S, James SJ, Hahn J. Mathematical modeling of the methionine cycle and transsulfuration pathway in individuals with autism spectrum disorder. J Theor Biol 2017; 416:28–37. <u>Crossref</u>
- 62. Wang H, Liang S, Wang M, et al. Potential serum biomarkers from a metabolomics study of autism. J Psychiatry Neurosci 2016;41:27–37. <u>Crossref</u>
- 63. West PR, Amaral DG, Bais P, et al. Metabolomics as a tool for discovery of biomarkers of autism spectrum disorder in the blood plasma of children. PLoS One 2014;9:e112445. <u>Crossref</u>
- 64. Xiong X, Liu D, Wang Y, Zeng T, Peng Y. Urinary 3-(3-hydroxyphenyl)-3-hydroxypropionic acid, 3hydroxyphenylacetic acid, and 3-hydroxyhippuric acid are elevated in children with autism spectrum disorders. Biomed Res Int 2016;2016:9485412. Crossref
- 65. Yan CL, Zhang J, Hou Y. Decreased plasma levels of lipoxin A4 in children with autism spectrum disorders. Neuroreport 2015;26:341–5. <u>Crossref</u>
- Al-Ayadhi LY, Ben Bacha AG, Kotb M, El-Ansary AK. A novel study on amyloid beta peptide 40, 42 and 40/42 ratio in Saudi autistics. Behav Brain Funct 2012;8:4. <u>Crossref</u>
- 67. Cai J, Ding L, Zhang JS, Xue J, Wang LZ. Elevated plasma levels of glutamate in children with autism spectrum disorders. Neuroreport 2016;27:272–6. <u>Crossref</u>
- 68. Chen J, Xin K, Wei J, Zhang K, Xiao H. Lower maternal serum 25(OH) D in first trimester associated with higher autism risk in Chinese offspring. J Psychosom Res 2016;89:98–101. <u>Crossref</u>

- 69. El-Ansary AK, Ben Bacha AG, Al-Ayadhi LY. Proinflammatory and proapoptotic markers in relation to mono and di-cations in plasma of autistic patients from Saudi Arabia. J Neuroinflammation 2011;8:142. <u>Crossref</u>
- 70. Pagan C, Delorme R, Callebert J, et al. The serotonin-N-acetylserotonin-melatonin pathway as a biomarker for autism spectrum disorders. Transl Psychiatry 2014;4:e479. <u>Crossref</u>
- Wang J, Zou Q, Han R, Li Y, Wang Y. Serum levels of glial fibrillary acidic protein in Chinese children with autism spectrum disorders. Int J Dev Neurosci 2017; 57:41–5. <u>Crossref</u>
- 72. Wang M, Chen H, Yu T, Cui G, Jiao A, Liang H. Increased serum levels of brain-derived neurotrophic factor in autism spectrum disorder. Neuroreport 2015;26:638–41. <u>Crossref</u>
- 73. Zhang QB, Gao SJ, Zhao HX. Thioredoxin: a novel, independent diagnosis marker in children with autism. Int J Dev Neurosci 2015;40:92–6. <u>Crossref</u>
- Zhang QB, Jiang LF, Kong LY, Lu YJ. Serum brainderived neurotrophic factor levels in Chinese children with autism spectrum disorders: a pilot study. Int J Dev Neurosci 2014;37:65–8. <u>Crossref</u>
- 75. Zhao HX, Yin SS, Fan JG. High plasma neopterin levels in Chinese children with autism spectrum disorders. Int J Dev Neurosci 2015;41:92–7. <u>Crossref</u>

- Chan AS, Sze SL, Cheung MC. Quantitative electroencephalographic profiles for children with autistic spectrum disorder. Neuropsychology 2007;21:74–81. Crossref
- 77. Duffy FH, Als H. A stable pattern of EEG spectral coherence distinguishes children with autism from neuro-typical controls a large case control study. BMC Med 2012;10:64. <u>Crossref</u>
- Eldridge J, Lane AE, Belkin M, Dennis S. Robust features for the automatic identification of autism spectrum disorder in children. J Neurodev Disord 2014;6:12. <u>Crossref</u>
- 79. Jamal W, Das S, Oprescu IA, Maharatna K, Apicella F, Sicca F. Classification of autism spectrum disorder using supervised learning of brain connectivity measures extracted from synchrostates. J Neural Eng 2014;11:046019. <u>Crossref</u>
- 80. Karhson DS, Golob EJ. Atypical sensory reactivity influences auditory attentional control in adults with autism spectrum disorders. Autism Res 2016;9:1079–92. <u>Crossref</u>
- 81. Matlis S, Boric K, Chu CJ, Kramer MA. Robust disruptions in electroencephalogram cortical oscillations and large-scale functional networks in autism. BMC Neurol 2015;15:97. <u>Crossref</u>